

STANDARD OPERATING PROCEDURE

Dishwashing Protocol

Zhou Lab, Institute for Environmental Genomics

Joy Van Nostrand, Lab Manager

2030 SRTC, 5-4403

Jan 11, 2019

Minimum Personal Protective Equipment Required: Glove, Lab coat, Long pants, Closed toed shoes

Special Notes:

****It is the responsibility of each individual person to clean up after themselves and to make sure their glassware, plasticware and other reusable items are cleaned and put away****

We have undergrad lab assistants who can help with cleaning, but they will not be doing all of the cleaning. Do **not** leave any glassware around the sinks unless you have:

- 1 – Autoclaved items containing cultures or antibiotics
- 2 – Removed stoppers, caps, and seals
- 3 - Poured out any liquid or other contents
- 4 – Removed tape or marker
- 5 – Rinsed out the container and removed any remaining residue
- 6 – Placed anaerobic tubes or serum bottles into an acid bath and labeled with date and time (make sure you have a label indicating it is an acid bath)

Protocol/Procedure:

General Glassware

1. Autoclave glassware if necessary – dump liquid agar into trash can
 - a. Items that should be autoclaved include, mortars and pestles, any cultures, media or buffers containing antibiotics, anything that has come in contact with cultures or cells
2. Discard any hazardous waste into the appropriate container (ask lab manager if you have any questions about where you should place the contents)
3. Pour out any buffer or media into the “used media” beaker in the hood in 2040 (discard down sink after 24 hr)
4. Remove tape (can use a razor blade if necessary) and marker (acetone will remove marker)
5. For anaerobic tubes and serum bottles, place into an acid bath for 24 h (instructions next page)
6. Scrub out any bottles with a bottle brush to remove any residue

7. Wash dishes with dishwasher or by hand
 - a. Load dishwasher, fill dispenser with dishwasher soap, start (use high temp, scrub, heat dry settings) – mortars and pestles should be washed in dishwasher
 - b. Put your name and date/time you are starting the dishwasher on the dry erase board on the dishwasher
 - c. If handwashing, fill a tub with hot water, add Alconox (powdered soap specifically designed for labware). Scrub out glassware with soapy water and bottle brush, rinse well with tap water and then distilled water, hang on rack to dry
8. Put away clean, dry labware

FEP Oak Ridge Tubes

1. Remove labels or marker
2. Wash in dishwasher or by hand as described above
3. Boil for 30 min in DI water
4. Hang on pegs to dry
5. DO NOT AUTOCLAVE

Acid bath

1. Look over *Acids SOP* before starting to prepare an acid bath
2. Wear disposable neoprene gloves (above the acid bath area)
3. Always set up and store the acid baths on the counter, NEVER the floor
4. Place tubes in a large autoclave bin
5. Fill bin with enough water to cover the tubes
6. Add ~90 mL sulfuric acid to the bath. Pour sulfuric acid into the beaker in the fume hood and then take the beaker to the bin of water
9. Label “Acid Bath” and date
10. If reusing the acid baths, they should be discarded each week

Disposing of bath

1. Add baking soda to neutralize (shake the baking soda over the bin, about 1/5 of a box to start)
2. Check pH with pH strips
3. Add more baking soda until pH reaches 7
4. Bath can then be poured down the sink