

Table S1. Primers and PCR conditions used in this paper

Primers	Sequence (5' – 3')	Thermal conditions	Reference
Rhizobacteria			
27F	AAC MGG ATT AGA TAC CCK G	94°C, 5 min, 1 cycle	Lane, 1990
1492R	GGY TAC CTT GTT ACG ACT T	94°C for 1 min, 48-58°C (gradient = 48.0, 48.3, 48.9, 49.7, 113 50.8, 52.3, 54.0, 55.4, 56.5, 57.3, 57.8, 58.0°C) for 45 s, 72°C for 1 min, 30 cycles 72°C for 8 min, 1 cycle	
Detection of 'Candidatus Liberibacter asiaticus'			
Conventional PCR			
A2	TAT AAA GGT TGA CCT TTC GAG TTT	94°C for 2 min, 1 cycle,	Hocquellet et al, 1999
J5	ACA AAA GCA GAA ATA GCA CGA ACA A	94°C for 10 s, 65°C for 10s, 72°C for 1 min, 35 cycles 72°C for 4 min, 1 cycle	
qPCR			
CQULA04F	TGG AGG TGT AAA AGT TGC CAA A	50°C for 2 min, 1 cycle	Wang et al., 2006
CQULAP10P	ATC GTC TCG TCA AGA TTG CTA TCC GTG ATA CTA G	95°C for 15 min, 1 cycle	
CQULA04R	CCA ACG AAA AGA TCA GAT ATT CCT CTA	94°C for 15 s and 60°C for 1 min, 45 cycles	
Taxon/Group specific qPCR			
Total bacteria			
Eub338	ACT CCT ACG GGA GGC AGC AG	95°C, 15 min, 1 cycle	Fierer et al., 2005
Eub518	ATT ACC GCG GCT GCT GG	95°C for 1 min, 53°C for 30 s, 72°C for 1 min, 40 cycles	
Acidobacteria			
Acid31	GAT CCT GGC TCA GAA TC	95°C, 15 min, 1 cycle	Fierer et al., 2005
Eub518	ATT ACC GCG GCT GCT GG	95°C for 1 min, 50°C for 30 s, 72°C for 1 min, 40 cycles	
Actinobacteria			
Actino235	CGC GGC CTA TCA GCT TGT TG	95°C, 15 min, 1 cycle	Fierer et al., 2005
Eub518	ATT ACC GCG GCT GCT GG	95°C for 1 min, 60°C for 30 s, 72°C for 1 min, 40 cycles	
Alphaproteobacteria			
Eub338	ACT CCT ACG GGA GGC AGC AG	95°C, 15 min, 1 cycle	Fierer et al., 2005
Alfa685	TCT ACG RAT TTC ACC YC TAC	95°C for 1 min, 60°C for 30 s, 72°C for 1 min, 40 cycles	

<i>Bacteroidetes</i>		95°C, 15 min, 1 cycle	Fierer <i>et al.</i> , 2005
Cfb319	GTA CTG AGA CAC GGA CCA	95°C for 1 min, 65°C for 30 s, 72°C for 1 min, 40 cycles	
Eub518	ATT ACC GCG GCT GCT GG		
<i>Betaproteobacteria</i>		95°C, 15 min, 1 cycle	Fierer <i>et al.</i> , 2005
Eub338	ACT CCT ACG GGA GGC AGC AG	95°C for 1 min, 60°C for 30 s, 72°C for 1 min, 40 cycles	
Bet680	TCA CTG CTA CAC GYG		
<i>Firmicutes</i>		95°C, 15 min, 1 cycle	Fierer <i>et al.</i> , 2005
Lgc353	GCA GTA GGG AAT CTT CCG	95°C for 1 min, 60°C for 30 s, 72°C for 1 min, 40 cycles	
Eub518	ATT ACC GCG GCT GCT GG		
<i>Pseudomonas spp.</i>		95°C, 15 min, 1 cycle	Drigo <i>et al.</i> , 2009
PsF	TTA GCT CCA CCT CGC GGC	95°C for 1 min, 64°C for 30 s, 72°C for 1 min, 40 cycles	
PsR	GGT CTG AGA GGA TGA TCA GT		
<i>Burkholderia spp.</i>		95°C, 15 min, 1 cycle	Drigo <i>et al.</i> , 2009
Burk3	CTG CGA AAG CCG GAT	95°C for 1 min, 64°C for 30 s, 72°C for 1 min, 40 cycles	
BurkR	TGC CAT ACT CTA GCY YGC		
<i>Bacillus spp.</i>		95°C, 15 min, 1 cycle	Drigo <i>et al.</i> , 2009
BacF	GGG AAA CCG GGG CTA ATA CCG GAT	95°C for 1 min, 63°C for 30 s, 72°C for 1 min, 40 cycles	
1378	CGG TGT GTA CAA GGC CCG GGA ACG		
qPCR of genes involved in nitrogen cycling			
<i>narG</i>		95°C, 15 min, 1 cycle	Hallin <i>et al.</i> , 2009
narG1960m2F	TAY GTS GGG CAG GAR AAA CTG	95°C for 15 s, 65 to 60°C for 30 s (-1°C by cycle), 72°C for 30 s, 80°C for 15 s, 6 cycles	
narG2050m2R	CGT AGA AGA AGC TGG TGC TGT T	95°C for 15 s, 60°C for 30 s, 72°C for 30 s, 80°C for 15 s, 40 cycles	
		95°C for 15 s, 60 to 95°C, 1 cycle	
<i>nirK</i>		95°C, 15 min, 1 cycle	Hallin <i>et al.</i> , 2009
nirK876	ATY GGC GGV CAY GGC GA	95°C for 15 s, 63 to 58°C for 30 s (-1°C by cycle), 72°C for 30 s, 80°C for 15 s, 6 cycles	
nirK1040	GCC TCG ATC AGR TTR TGG TT		

<i>nirS</i> nirSCd3aFm nirSR3cdm	AAC GYS AAG GAR ACS GG GAS TTC GGR TGS GTC TTS AYG AA	95°C for 15 s, 60°C for 30 s, 72°C for 30 s, 80°C for 15 s, 40 cycles 95°C for 15 s, 60 to 95°C, 1 cycle 95°C, 15 min, 1 cycle 95°C for 15 s, 65 to 60°C for 30 s (-1°C by cycle), 72°C for 30 s, 80°C for 15 s, 6 cycles 95°C for 15 s, 60°C for 30 s, 72°C for 30 s, 80°C for 15 s, 40 cycles 95°C for 15 s, 60 to 95°C, 1 cycle 95°C, 15 min, 1 cycle	Hallin <i>et al.</i> , 2009
<i>nosZ</i> nosZ2F nosZ2R´	CGC RAC GGC AAS AAG GTS MSS GT CAK RTG CAK SGC RTG GCA GAA	95°C for 15 s, 65 to 60°C for 30 s (-1°C by cycle), 72°C for 30 s, 80°C for 15 s, 6 cycles 95°C for 15 s, 60°C for 30 s, 72°C for 30 s, 80°C for 15 s, 40 cycles 95°C for 15 s, 60 to 95°C, 1 cycle 95°C, 10 min, 1 cycle	Hallin <i>et al.</i> , 2009
<i>amoA(AOB)</i> amoA-1F amoA-2R	GGG GTT TCT ACT GGT GGT CCC CTC KGS AAA GCC TTC TTC	94°C for 45 s, 58°C for 45 s, 72°C for 45 s, 39 cycles 95°C for 15 s, 60°C for 30 s, to 95°C for 15 s, 1 cycle	Hallin <i>et al.</i> , 2009
<i>amoA(AOA)</i> 19F CrenamoA616r48x	ATG GTC TGG CTW AGA CG GCC ATC CAB CKR TAN GTC CA	95°C, 10 min, 1 cycle 94°C for 45 s, 55°C for 45 s, 72°C for 45 s, 39 cycles 95°C for 15 s, 60°C for 30 s, to 95°C for 15 s, 1 cycle	Hallin <i>et al.</i> , 2009

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Figure S1. Rarefaction curves of 16S rRNA gene clone libraries from the rhizosphere of healthy citrus (HT1, HT2, and HT3); '*Ca. Liberibacter asiaticus*' infected citrus (IT1, IT2, and IT3); and bulk soil samples.

Figure S2. Relative abundances of the different bacterial phyla/genus in the rhizosphere of healthy (HT1, HT2, HT3); '*Ca. Liberibacter asiaticus*' infected citrus (IT1, IT2, and IT3); and bulk soil samples, as estimated using the qPCR assays. Error bars represent the standard errors of the means for three replicates. Statistical significance is represented by letters above each bar, with different letters signifying differences at $P \leq 0.01$ for each bacterial phyla/genus.

Figure S3. Hierarchical cluster analysis of (A) ribulose-1, 5-bisphosphate carboxylase / oxygenase (Rubisco); (B) carbon monoxide dehydrogenase (CODH); and (C) propionyl-CoA / acetyl-CoA carboxylase (PCC / ACC) genes in rhizosphere of '*Ca. L. asiaticus*' infected (IT1-IT3) or healthy (HT1-HT3) citrus. Genes that were present in at least three samples were used for cluster analysis. Results were generated in CLUSTER and visualized using TREEVIEW. Red indicates signal intensities above background while black indicates signal intensities below background. Brighter red colouring indicates higher signal intensities.

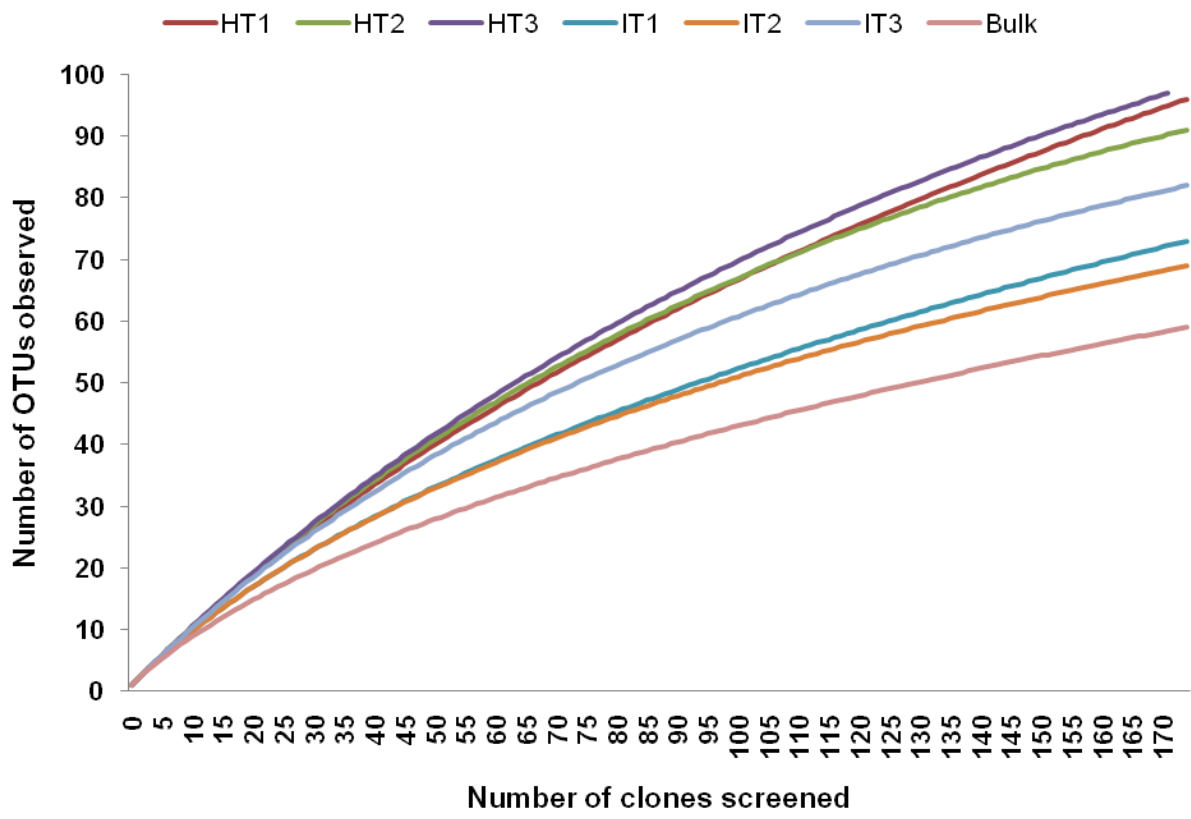


Fig. S1

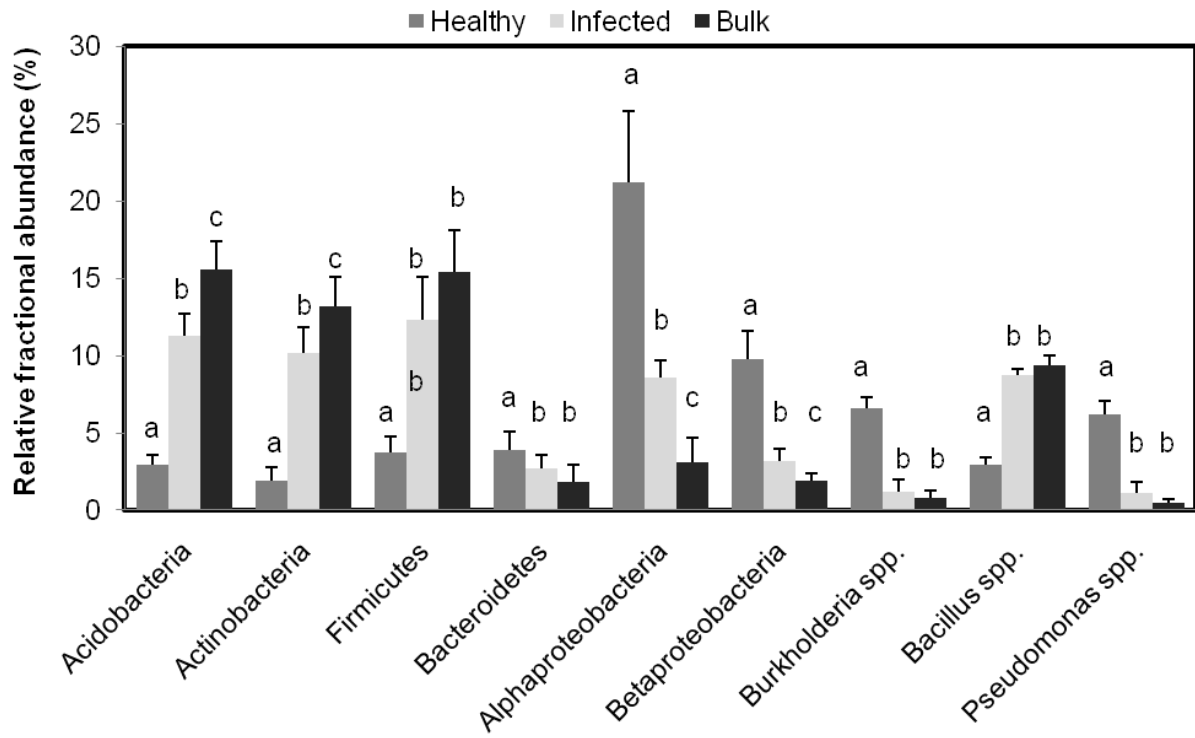


Fig. S2

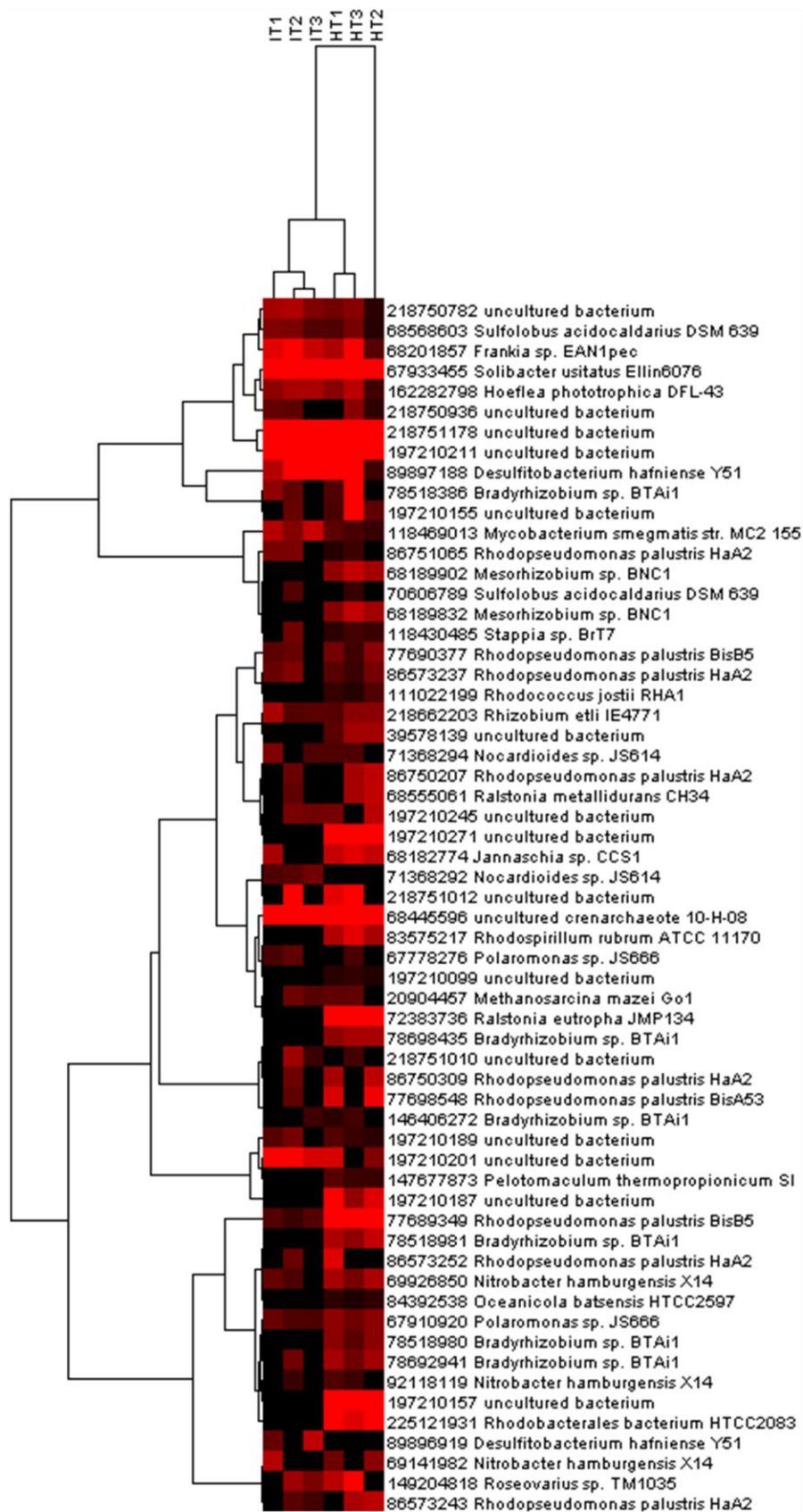


Fig. S3A

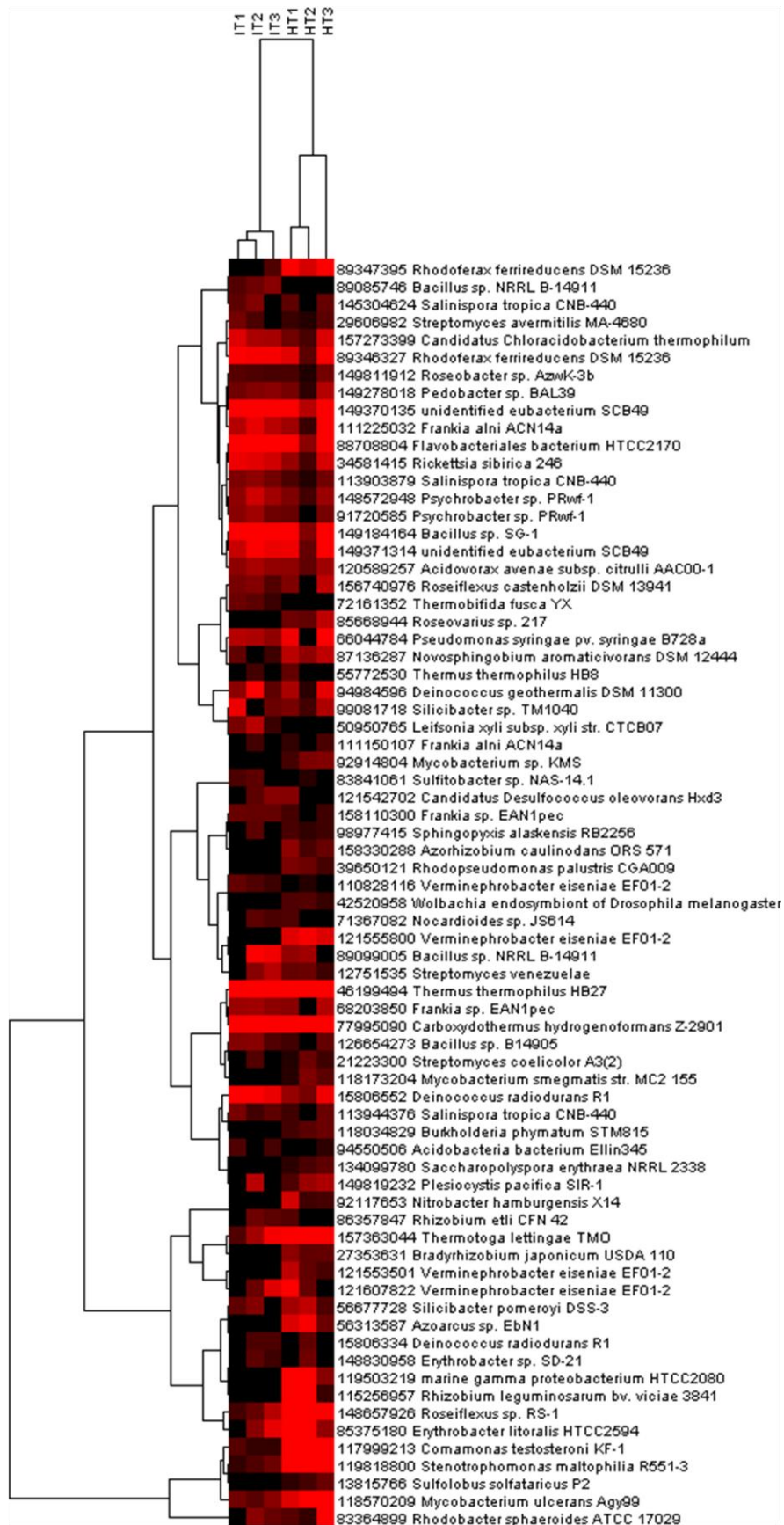


Fig. S3B

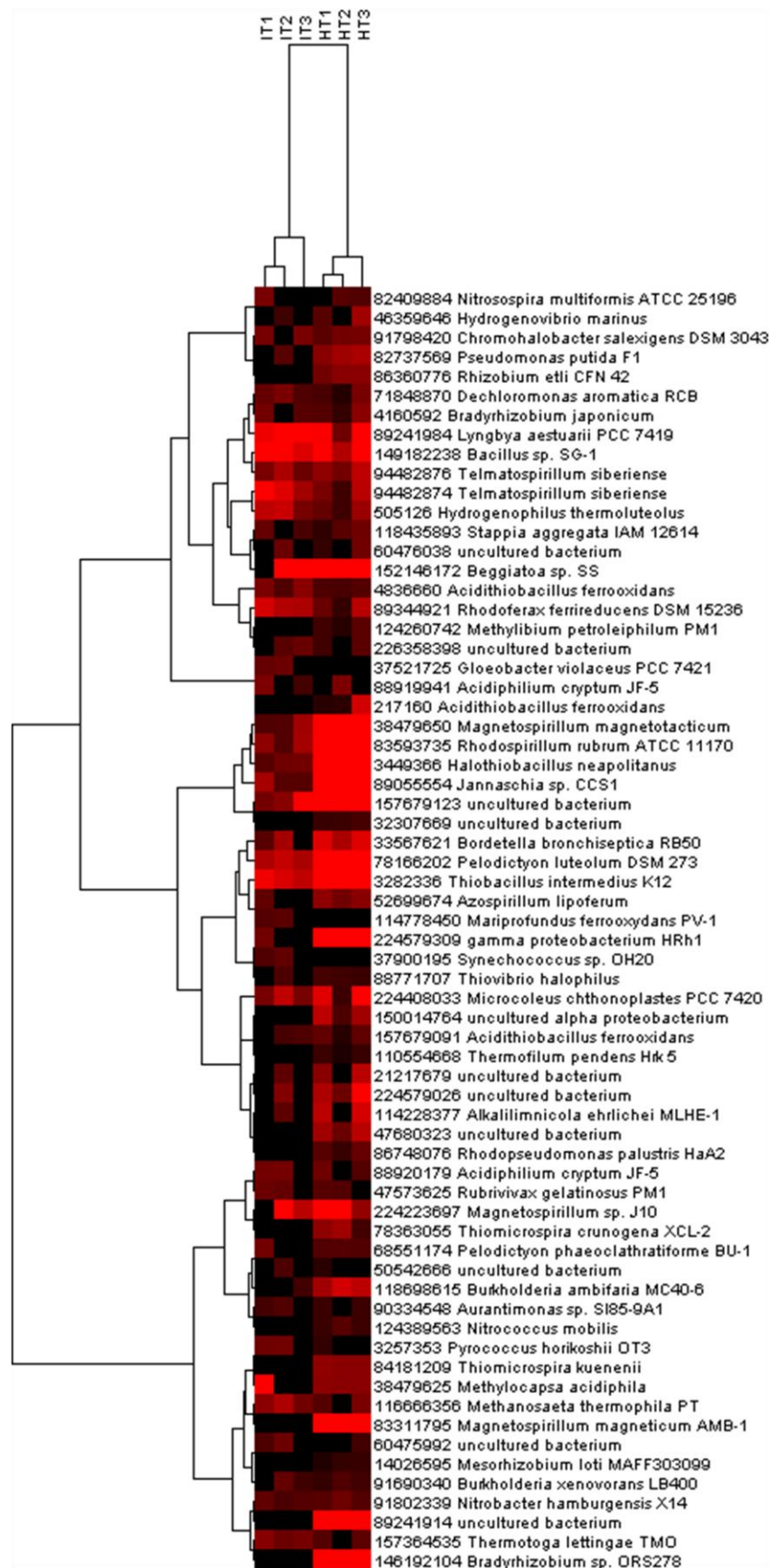


Fig. S3C