1	(Manuscript number: AEM07758-11)
2	Genetic linkage of soil carbon pools and microbial functions in
3	subtropical freshwater wetlands in response to experimental warming
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5	Supplementary Information
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21 A. MATERIALS AND METHODS

22 SI-1: Microcosm configuration

A microcosm setup (Fig. S1) for simulating climate warming at a minute-scale under 23 both daily and seasonal scenarios was developed for this study by using independently 24 monitored water-bath jackets. Microcosms consisted of four major components: a storage 25 section, a heating section, a water circulation section, and a real-time control section. The 26 storage section was composed of two stainless steel incubation boxes: one for the 27 present-day ambient temperature treatment (Control), and the other for the 28 +5°C-increased temperature treatment (Warmed). The real-time control section was 29 composed of a computer (HP a6315cn), a custom-built controller (TZ2008, Jiaxing 30 China), digital temperature probes (NB 407-25a, China), and lab-designed software (C⁺⁺ 31 language). The temperature probes, heater, and pump were programmed through the 32 custom-built controller by the computer. With the help of the software, the temperatures 33 in both incubation boxes were continuously recorded and differences in the box 34 temperatures were compared by digital probes at two-minute intervals. The temperature 35 difference between the two incubation boxes was set at 5°C±1°C. The custom-built 36 controller simultaneously turned the heater and pump on or off when the temperature 37 difference was less than 4°C or more than 6°C, respectively. Except for the computer and 38 the controller, the rest of the microcosm components were set up outdoors in May 2008. 39 40 This novel microcosm offers a high resolution temperature comparison, repeatability, and the capability for simulating more realistic warming conditions. 41

B. SUPPORTING TABLES

44	Wetland ID	County	Latitude and longitude	Main wetland use	Annual mean water depth, m	Annual mean flow rate, m min ⁻¹	Dominant macrophytes
	YaTang riverine wetland (YT)	TongXiang	120°29'13"E, 30°43'15"N	Mixed use	0.80	1.02	Trapa bispinosa, Alternanthera philoxeroides, Trapa spp, Arundo dona, Arundo donax
	XiaZhuhu (XZ)	DeQing	120°02'54"E, 30°31'28"N	Tourism and aquaculture	1.50	0.12	Trapa bispinosa, Alternanthera philoxeroides, Trapa spp, Arundo dona, Arundo donax
	XiXi national wetland park (XX)	HangZhou City	120°03'59"E, 30°16'23"N	Tourism	0.85	0.10	Phragmites communis, Trapa spp, Acorus calamus, Sagittaria sagittifolia, Miscanthus floridulus

43	Table S1	Description	and basic	properties	of wetland	sites fo	or this study
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Table S2 Overall microbial diversities and detected functional gene shifts by GeoChip analysis for samples collected from YT, XZ and XX
wetland columns in the microcosm experiment under warmed and control treatments (Control: ambient temperature; Warmed: ambient
temperature +5°C). Values in parentheses are percentages.

Wetland columns		Ge	enes	Simpson	Total no. of	
		Overlap	Uniqueness	Diversity Index $(1/D)$	Evenness (E_H)	genes detected
VT	, C	10960(67.2)	891 (6.98)	4218	0.33	12771
I I	W	10800 (07.3)	1454 (10.2)	3948	0.28	14216
V7	С	$((0,0))^{a}$	163 (2.17)	2010	0.27	7499
ΛL	W	0000 (34.8)	518 (4.64)	3380	0.30	11156
vv	С	9502(62,2)	415 (4.09)	2639	0.26	10147
$\Lambda\Lambda$	W	8502 (05.5)	731 (6.20)	2895	0.25	11785
<i>p</i> -value			0.033	0.449	0.707	0.086

Table S3 Top 20 genes with the highest normalized signal intensities involved in carbon degradation for starch, hemicellulose, cellulose, chitin and lignin degradation detected in warmed and control samples from tested wetland columns in the microcosm experiment (Control: ambient temperature; Warmed: ambient temperature + 5°C). All data are expressed as mean, with standard deviation in parentheses. The *p*-values are from Student's *t*-tests on the difference between warming and control treatments. Significant (< 0.05) and marginally significant (< 0.1) *p*-values

are in bold.

Gene information	Functional category	Control	Warmed	<i>p</i> -value
133778506 (uncultured bacterium)	Chitin	15.97 (8.24)	11.75 (3.93)	0.144
209503326 (Burkholderia sp. H160)	Cellulose	2.86 (0.95)	7.06 (1.55)	0.007
9105753 (Xylella fastidiosa 9a5c)	Chitin	7.94 (5.42)	6.66 (5.65)	0.074
40714547 (Trametes versicolor)	Lignin	3.23 (1.61)	6.38 (2.97)	0.047
262195882 (Haliangium ochraceum DSM 14365)	Starch	3.09 (2.10)	6.34 (3.66)	0.035
239930964 (Streptomyces ghanaensis ATCC 14672)	Chitin	2.32 (1.43)	5.65 (3.40)	0.075
256769453 (Streptomyces sp. C)	Lignin	1.71 (0.58)	5.40 (4.67)	0.130
29160339 (Botryosphaeria rhodina)	Cellulose	2.38 (0.92)	4.75 (1.18)	0.007
209516215 (Burkholderia sp. H160)	Cellulose	3.36 (1.34)	4.74 (0.14)	0.123
76876845 (Pseudoalteromonas haloplanktis TAC125)	Lignin	2.51 (0.23)	4.72 (4.43)	0.230
240278619 (Ajellomyces capsulatus H143)	Chitin	2.73 (0.96)	4.62 (2.70)	0.111
145304813 (Salinispora tropica CNB-440)	Cellulose	5.75 (1.54)	4.44 (0.29)	0.163
21929222 (Phanerochaete sordida)	Lignin	2.96 (1.80)	4.42 (2.16)	0.061
88814399 (marine actinobacterium PHSC20C1)	Hemicellulose	1.69 (0.58)	4.37 (1.61)	0.070
239982330 (Streptomyces albus J1074)	Chitin	3.72 (1.36)	4.23 (0.80)	0.217
256686144 (Jonesia denitrificans DSM 20603)	Chitin	2.16 (0.58)	4.01 (1.97)	0.160
115353435 (Chaetomium globosum)	Chitin	3.21 (2.02)	3.47 (2.85)	0.325
170654540 (Methylobacterium radiotolerans JCM 2831)	Starch	2.20 (1.44)	3.11 (1.80)	0.101
269923210 (Brevundimonas subvibrioides ATCC 15264)	Starch	1.33 (0.94)	3.04 (2.55)	0.119
239928121 (Streptomyces ghanaensis ATCC 14672)	Cellulose	2.11 (2.02)	2.98 (1.04)	0.267





Fig. S1 Schematic of the experimental wetland microcosm system developed using
independently monitored water-bath jackets under the current climate condition (Left:
ambient temperature, control) and the simulated climate warming condition (Right:
ambient temperature + 5°C, warmed).





Fig. S2 Linear correlations (p < 0.01) between microbial biomass carbon (MBC) and soil organic carbon pools, including total organic carbon (TOC) and labile organic carbon pools (HLOC, MLOC, and LOC) for tested subtropical wetland soils in the microcosm experiment. The linear regression model for MBC and TOC is y = 36.4x -27.4, adj. $R^2 = 0.474$; MBC and LOC: y = 21.1x - 15.9, adj. $R^2 = 0.471$; MBC and MLOC: y = 18.9x - 15.0, adj. $R^2 = 0.623$; MBC and HLOC: y = 10.9x - 7.7, adj. $R^2 =$ 0.663.



Fig. S3 Microbial community structure features of main phyla for (A) bacteria and (B) fungi and archaea as indicated by detected gene numbers under warmed and control treatments (Control: ambient temperature; Warmed: ambient temperature $+5^{\circ}$ C). α -, β -, γ-, δ-Proteobacteria, Actinobacteria, Firmicutes and Cyanobacteria are phyla of bacteria; Ascomycota and Basidiomycota are phyla of fungi, and Crenarchaeota and Euryarchaeota are phyla of archaea.



Sequences detected only in warmed soil

Fig. S4 Gene sequences responsible for lignin degradation involved in carbon cycling were detected in warmed samples but were absent in control samples. The signal intensity for each sequence was the average of the total signal intensity from all warmed samples. Error bars are + 1 standard deviation. Gene number on the x-axis is the protein ID number for each gene as listed in the GenBank database.