

ORIGINAL ARTICLE

Reproducibility and quantitation of amplicon sequencing-based detection

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To determine the reproducibility and quantitation of the amplicon sequencing-based detection approach for analyzing microbial community structure, a total of 24 microbial communities from a long-term global change experimental site were examined. Genomic DNA obtained from each community was used to amplify 16S rRNA genes with two or three barcode tags as technical replicates in the presence of a small quantity (0.1% wt/wt) of genomic DNA from *Shewanella oneidensis* MR-1 as the control. The technical reproducibility of the amplicon sequencing-based detection approach is quite low, with an average operational taxonomic unit (OTU) overlap of $17.2\% \pm 2.3\%$ between two technical replicates, and $8.2\% \pm 2.3\%$ among three technical replicates, which is most likely due to problems associated with random sampling processes. Such variations in technical replicates could have substantial effects on estimating β -diversity but less on α -diversity. A high variation was also observed in the control across different samples (for example, 66.7-fold for the forward primer), suggesting that the amplicon sequencing-based detection approach could not be quantitative. In addition, various strategies were examined to improve the comparability of amplicon sequencing data, such as increasing biological replicates, and removing singleton sequences and less-representative OTUs across biological replicates. Finally, as expected, various statistical analyses with preprocessed experimental data revealed clear differences in the composition and structure of microbial communities between warming and non-warming, or between clipping and non-clipping. Taken together, these results suggest that amplicon sequencing-based detection is useful in analyzing microbial community structure even though it is not reproducible and quantitative. However, great caution should be taken in experimental design and data interpretation when the amplicon sequencing-based detection approach is used for quantitative analysis of the β -diversity of microbial communities.

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Introduction

Soil microbial communities are among the most complex, diverse and important assemblages in the biosphere. It is estimated that 1 g of forest soil contains 4×10^7 prokaryotic cells, whereas 1 g of grassland soil has 2×10^9 prokaryotic cells (Daniel, 2005). DNA reassociation kinetic studies have estimated that 1 g of soil contains approximately 2000–18 000 genomes (Torsvik *et al.*, 1996, 1998; Torsvik and Ovreas, 2002), suggesting that

prokaryotic diversity in soil is extremely high. Owing to such high diversity, soil microbial communities are among the most difficult to characterize both phenotypically and genetically. Although introduction of molecular biology approaches have greatly advanced microbial community studies (Zhou *et al.*, 2004), our understanding of the extent of microbial diversity in soil environments is still very limited, especially in contrast to plants and animals. In addition, due to the uncultivated status of the majority of microorganisms (>99%) in nature, understanding their genetic properties, metabolic characteristics, biochemical functions and ecological functions is even more difficult.

Large-scale genome sequencing and associated genomic technologies have revolutionized the study of microbial communities. Recently, several new high-throughput sequencing approaches such as 454 pyrosequencing, Illumina (Illumina, San Diego,

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CA, USA) and SOLiD (Applied Biosystems, Foster City, CA, USA) have been developed (Margulies *et al.*, 2005; Hamady *et al.*, 2008; Smith *et al.*, 2008; Schulte *et al.*, 2010). At present, the 454 pyrosequencing technology has been widely used to reveal microbial diversity and ecology in different natural environments, such as the deep sea (Sogin *et al.*, 2006; Huber *et al.*, 2007), extreme hydrogeological conditions (Edwards *et al.*, 2006) and soil (Leininger *et al.*, 2006; Roesch *et al.*, 2007). In addition, multiplexed high-throughput pyrosequencing of individual genes (for example, 16S rRNA) by tagging or bar coding with short nucleotides (also called pyrotag sequencing (Kunin *et al.*, 2010)) (Huse *et al.*, 2007; Parameswaran *et al.*, 2007; Roesch *et al.*, 2007; Hamady *et al.*, 2008) has been developed to process many samples simultaneously and has been widely used in microbial community studies (Youssef *et al.*, 2009; Cheung *et al.*, 2010; Chun *et al.*, 2010; He *et al.*, 2010a,b; Koopman *et al.*, 2010; Schutte *et al.*, 2010; Teixeira *et al.*, 2010; Uroz *et al.*, 2010). A similar strategy has also been used for sequencing functional genes (Iwai *et al.*, 2010). Thus, as PCR-amplified products (amplicons) are used for sequencing, this approach is generally referred to as amplicon sequencing. Various studies have clearly demonstrated that amplicon sequencing is powerful for studying microbial community diversity and activity.

Some technical problems inherent to amplicon sequencing have recently been observed, such as overestimating the rare biosphere because of pyrosequencing errors (Quince *et al.*, 2008, 2009; Kunin *et al.*, 2010) and overestimating gene and taxon abundance because of inherent, systematic artifacts (Gomez-Alvarez *et al.*, 2009). Amplicon size and primer pair also seem to have a great influence on estimating microbial community richness and evenness (Engelbrektson *et al.*, 2010). However, it is unclear whether amplicon sequencing-based detection is reproducible and quantitative. In this study, we have examined these important issues by analyzing the composition and structure of 24 microbial communities from a field site in Oklahoma (USA) for a long-term study of global climate change. Genomic DNA obtained from each community was used to amplify 16S rRNA genes with two or three bar-coded tags as technical replicates in the presence of a small quantity (0.1% wt/wt) of *Shewanella oneidensis* MR-1 genomic DNA as the control. In this study, technical replicates mean that the same genomic DNA obtained from a community is amplified multiple times and subsequently sequenced. Our results indicated that the reproducibility among technical replicates and quantitative capability were quite low for amplicon sequencing-based detection. Various strategies for improving data comparability are discussed. Although experimental data are generated by the pyrosequencing approach in this study, the conclusions should be applicable to other sequencing approaches as well.

Materials and methods

Site description and sampling

Soil samples used in this study were obtained from the KFFL (Kessler Farm Field Laboratory) at the Great Plain Apiaries in McClain County (OK, USA; 34°58'54"N, 97°31'14"W) and were part of a long-term experiment using a paired nested design with warming as the main factor and clipping as a secondary factor (Luo *et al.*, 2001). There were six replicates for each of the following treatments: (1) clipped, warmed, (2) unclipped, warmed, (3) clipped control and (4) unclipped control. The warming plots have been continually warmed for ~2 °C since November 1999. A total of 24 soil samples were taken from a 0–15 cm layer in April 2007. Each sample was composited from four soil cores (2 cm diameter × 15 cm deep) after being sieved (2 mm). All samples were stored at –80 °C.

DNA extraction

Soil DNA was extracted by freeze-grinding mechanical lysis as described previously (Zhou *et al.*, 1996) and was purified using a low melting agarose gel, followed by phenol extraction. DNA quality was assessed based on the absorbance ratios of 260/280 nm and 260/230 nm using NanoDrop ND-1000 Spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA), whereas DNA concentration was quantified by PicoGreen (Promega, Sunnyvale, CA, USA) (Ahn *et al.*, 1996) using FLUOstar Optima (BMG Labtech, Jena, Germany).

Sample tagging and PCR amplicon preparations for sequencing

On the basis of the V4–V5 hypervariable regions of bacterial 16S rRNAs (*Escherichia coli* positions 515–907), the PCR primers, F515: GTGCCAGCMGC CGCGG and R907: CCGTCAATTCMTTTRAGTTT were selected. Both primers covered >98% of the 16S gene sequences in the ribosomal database (Cole *et al.*, 2007, 2009). To pool multiple samples for one run of 454 sequencing, a sample tagging approach was used (Binladen *et al.*, 2007; Hamady *et al.*, 2008). Each tag (6 mer) was added to the 5'-end of both forward and reverse primers (Supplementary Table S1), synthesized by Invitrogen (Carlsbad, CA, USA) and then used for the generation of PCR amplicons. To evaluate the reproducibility of sequencing results, a total of 60 sequencing tags were used: 12 samples were amplified with 3 tagged primers ($n=36$) and 12 with 2 tagged primers ($n=24$). Pyrosequencing data for four samples were excluded from data analysis because of some errors in primers used.

The amplification mix contained 10 Units of Pfu polymerase (BioVision, Mountain View, CA, USA), 5 µl Pfu reaction buffer, 200 µM dNTPs (Amersham, Piscataway, NJ, USA) and a 0.2 µM concentration of each primer in a volume of 50 µl. Genomic DNA

(10 ng) was added to each amplification mix. Cycling conditions were an initial denaturation at 94 °C for 3 min, 25 cycles of 95 °C for 30 s, 58 °C for 60 s, 72 °C for 60 s and a final 2-min extension at 72 °C. To obtain enough PCR products for sequencing, multiple (5–10) 100- μ l reactions were used for each sample. The products from each tagged primer were pooled and purified by agarose gel electrophoresis. The amplified PCR products were recovered and then quantitated using PicoGreen (Ahn *et al.*, 1996). Finally, amplicons of all samples were pooled in an equimolar concentration for 454 pyrosequencing.

454 Pyrosequencing

Pyrosequencing was carried out at the Genome Sequencing Center of the University of Oklahoma. In brief, fragments in the amplicon libraries were repaired and ligated to the 454 sequencing adapters, and the resulting products were bound to beads under conditions that favor one fragment per bead. The beads were emulsified in a PCR mixture in oil, and PCR amplification occurred in each droplet, generating millions of copies of a unique DNA template. After breaking the emulsion, DNA strands were denatured, and beads carrying single-stranded DNA clones were deposited into wells on a PicoTiter Plate (454 Life Sciences, Branford, CT, USA) for pyrosequencing (Margulies *et al.*, 2005) on a FLX 454 system. For this study, we recovered both forward and reverse sequencing reads of the 24 samples with an average length of ~240 bp. All pyrosequencing reads were initially processed using the RDP (Ribosomal Database Project) pyrosequencing pipeline (<http://pyro.cme.msu.edu/>) (Cole *et al.*, 2009).

Pyrosequencing data preprocessing

To minimize the effects of random sequencing errors, we eliminated low-quality sequences as follows: (1) sequences that did not perfectly match the PCR primer at the beginning of a read; (2) sequences with non-assigned tags; (3) short sequence reads (<200 bp) after the proximal PCR primer if they terminated before reaching the distal primer and (4) sequences that contained more than one undetermined nucleotide (N). Only the first 240 bp after the proximal PCR primer of each sequence was included as the quality of sequences degrades beyond this point. After removing low-quality sequences, the raw sequences were sorted and distinguished by unique sample tags and each sample had two or three unique tags as replicates. The tags and primers were then trimmed for each replicate.

On the basis of sequences obtained, there is ~120 bp overlap between the sequences from the forward and reverse primers. To identify sequences that potentially originated from the same 16S rRNA gene molecule, sequences from forward and reverse

primers were compared. If the number of overlapped nucleotides was >120 bp and their sequence identity was >97%, these two forward and reverse sequences were considered to originate from the same DNA molecule. If one sequence from the forward (or reverse) primer was >97% identical to multiple sequences, the one with the highest identity from the reverse (forward) primer was considered to be its corresponding counterpart. The two matched sequences were then combined to generate a longer sequence (356 nt). All combined sequences and individual sequences from both the forward and the reverse primers were then pooled as a full set of sequences of each tag.

All sequences from the 24 samples were aligned by RDP Infernal Aligner, a fast secondary-structure aware aligner (Ribosomal Database Project, Michigan State University, East Lansing, MI, USA) (Nawrocki and Eddy, 2007) and then complete linkage clustering was used to define operational taxonomic units (OTUs) within a 0.03 difference (Stackebrandt and Goebel, 1994). As 97% identity is used as the cutoff and singleton sequence reads were removed, each OTU contains more than two sequence reads from at least two samples. The sequence reads within an OTU were partitioned into individual samples based on the original sample-tagging information. In addition, sequences were then assigned to a taxonomy by the RDP classifier (Wang *et al.*, 2007) with a confidence cutoff of 0.8. The lineage of each OTU was summarized with all phylogenetic information.

As the sequence numbers of individual OTUs obtained varied significantly among different samples, the relative proportions of sequence numbers were used for subsequent analysis. The RA (relative abundance) was calculated as:

$$RA_{ij} = \frac{S_{ij}}{\sum_{j=1}^n S_{ij}},$$

where S_{ij} is the number of sequencing reads of the j th OTU (1 to n) in the i th sample (1 to m).

Statistical analysis

The Mothur program (University of Michigan Health System, Ann Arbor, MI, USA) (Patrick Schloss, <http://www.mothur.org/>) was used for rarefaction analysis at different levels based on individual tags, samples, treatments and the whole data set. Rarefaction curves were created by plotting the results of random sequence re-sampling from OTUs. The rarefaction curves were fitted with the model $y = a(1 - e^{-bx})$, and maximum OTUs were estimated as predicted diversity based on the fitted rarefaction curve. OTU predictions were also obtained using the Chao1 method (Chao, 1987). Both Sørensen similarity (S_s) and Bray-Curtis similarity (BC_s) were calculated between any pair of two samples, and

the complement (S_d) of Sørensen similarity ($1 - S_s$) and the complement (BC_d) of Bray-Curtis similarity ($1 - BC_s$) were used to measure β -diversity of microbial communities among different samples. One-way ANOVA (analysis of variance) was used to compare the β -diversity at technical (tag) replicate and biological replicate levels, and that between control and the treatments (warming and control). The Duncan multiple range test was used to determine statistical significance of the differences between microbial communities at different levels.

Ordination analyses were further performed using PC-ORD (McCune and Mefford, 1999) and confirmed by CANOCO 4.5 (Biometris—Plant Research International, Wageningen, The Netherlands). Detrended correspondence analysis was used to determine the overall phylogenetic composition changes in microbial communities. Detrended correspondence analysis is an ordination technique that uses detrending to remove the arch effect, in which data points are organized in a horseshoe-like shape, in correspondence analysis (Hill and Gauch, 1980).

Data sets of microbial communities generated by different analytical methods were used to examine whether warming or clipping had significant effects on soil microbial communities. Typically, it is difficult for all data sets to meet the assumptions (for example, normality, equal variances, independence) of parametric statistics. Thus, in this study, three different complementary non-parametric analyses for multivariate data were used: analysis of similarity (ANOSIM) (Clarke, 1993), non-parametric multivariate ANOVA (Adonis) using distance matrices (Anderson, 2001) and multiresponse permutation procedure (MRPP). We used the Bray-Curtis similarity index to calculate the distance matrix for ANOSIM, Adonis and MRPP analyses. MRPP is a non-parametric procedure that does not depend on assumptions, such as normally distributed data or homogeneous variances. Rather, it depends on the internal variability of data (Mielke and Berry, 2001; McCune and Grace, 2002). All three methods are based on dissimilarities among samples and their rank order to calculate test statistics. The Monte Carlo permutation was used to test the significance of statistics. All three procedures (ANOSIM, Adonis and MRPP) were performed using the Vegan package (version 1.15-1, Department of Statistics, Iowa State University, Ames, IA, USA) in R v. 2.8.1 (R Development Core Team, 2006).

Results

OTU overlaps among technical replicates

The V4-V5 regions of 16S rRNA genes were amplified and sequenced with one pyrosequencing run, and a total of 52.2 Mbp sequences with 213 329 reads were obtained (see Supplementary text for overview of sequencing statistics). A total of 115 741 sequence reads were obtained after preprocessing.

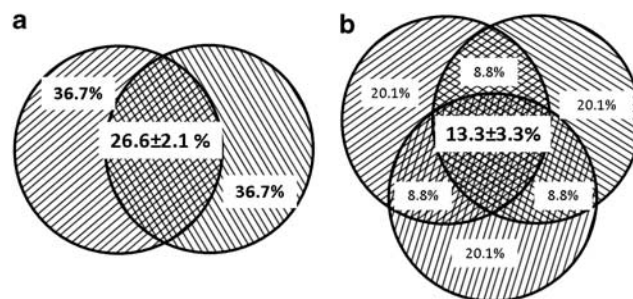


Figure 1 The average overlap of OTUs of microbial communities sequenced with two or three pyrotags. (a) Reverse primer sequences with two tags, singleton sequences removed. (b) Reverse primer sequences with three tags, singleton sequences removed.

An average of 1121 ± 390 OTUs were obtained for each tag based on the combined samples. Rarefaction analysis indicated that the maximum number of OTUs in this warming site is $\sim 17\,370$, and that the diversity of the abundant populations in these communities was recovered in this study (Supplementary text).

As community DNA from the same sample was amplified with two or three tagged primers as technical replicates, followed by sequencing, theoretically, a maximum of 100% overlap of OTUs detected among these technical replicates should have been expected if all populations in these communities were sampled. However, in reality, the expected percentage of overlap will be lower because of undersampling, random sampling and the complexity of microbial communities. Surprisingly, the overlap among these technical replicates was quite low (Supplementary Table S2). The average OTU overlap among the samples with two technical replicates for the forward primer was $15.5\% \pm 2.5\%$ (Supplementary Figure S1A), ranging from 10.9% to 19.2% (Supplementary Table S2A) and $18.8\% \pm 2.1\%$ (Supplementary Figure S1B) for the reverse primer with the range of 13.9–21.4% (Supplementary Table S2B). The difference between forward and reverse primers was significant ($P = 0.003$ based on Mann–Whitney U -test). The average OTU overlap among the samples with three technical replicates was $7.4\% \pm 2.1\%$ (Supplementary Figure S2A) in the range of 4.1–11.0% (Supplementary Table S2A) for the forward primer, and $9.1\% \pm 2.5\%$ (Supplementary Figure S2B) in the range of 3.5–12% for the reverse primer (Supplementary Table S2B). The difference between forward and reverse primers was also significant ($P = 0.05$ based on Mann–Whitney U -test). These results suggest that variations in technical replicates for the amplicon sequencing-based detection approach are quite high, which are most likely due to problems associated with random sampling processes.

The OTU overlap among technical replicates was increased significantly by removing singleton sequences. After removing singleton sequences, the

Table 1 One-way ANOVA and Duncan grouping method to assess β -diversity at the levels of technical replicates, biological replicates and treatments based on the combined OTUs from both forward and reverse primers

	Data size ^a	With singletons				Singletons removed			
		Sørensen		Bray-Curtis		Sørensen		Bray-Curtis	
		β -Diversity ^b	Significance ^c	β -Diversity ^b	Significance ^c	β -Diversity ^b	Significance ^c	β -Diversity ^b	Significance ^c
Technical replicate	42	0.756	c	0.763	c	0.734	c	0.475	b
Biological replicate	322	0.814	b	0.866	a	0.798	b	0.533	a
Warming	196	0.83	a	0.854	b	0.815	a	0.526	a
Clipping	196	0.834	a	0.853	b	0.82	a	0.525	a

Abbreviation: ANOVA, analysis of variance.

^aData sizes (n) are the number of data points of the pairwise comparisons within the technical replicates, biological replicates or treatments.

^bWe calculate two popular β -diversity dissimilarity measurements, Sørensen and Bray-Curtis, in which Sørensen dissimilarity is based on OTUs richness and Bray-Curtis dissimilarity takes OTUs abundance into account.

^cSignificance at $[pr(>F)] < 0.05$, using Duncan grouping method is shown. a, b and c represent the significance of β -diversity differences between technical replicates, biological replicates and treatments. a marks the highest β -diversity, the one less than the highest but not significant is still marked with a, then the ones significantly lower than the highest is marked with b or c.

OTU overlap among the samples with two technical replicates ranged from 15.1–30.2% (Supplementary Table S2A), with an average of $23.3\% \pm 4.3\%$ (Supplementary Figure S1C, $P < 0.001$ based on Mann–Whitney U -test) for the forward primer, and from 23.8% to 32.2% (Supplementary Table S2B) with an average of $26.6\% \pm 2.1\%$ (Figure 1a, $P < 0.001$) for the reverse primer. For samples with three technical replicates, they ranged from 6.2% to 16.6% (Supplementary Table S2A), with an average of $11.7\% \pm 3.3\%$ (Supplementary Figure S2C, $P = 0.009$) for the forward primer, and from 5.8% to 17.7% (Supplementary Table S2B) with an average of $13.3\% \pm 3.3\%$ (Figure 1b, $P = 0.005$) for the reverse primer. In addition, when biological replicates are available, one could remove the less-representative OTUs for a reliable diversity comparison across different samples. The percentages of OTUs overlap were significantly increased among technical replicates (Supplementary Table S2A–C) by removing the less-representative OTUs across all biological replicates within a treatment. In addition, to determine whether more abundant OTUs are reproducibly detected among technical replicates, OTUs were ranked based on their RA in each sample against their frequencies detected among the technical replicates. Interestingly, no consistent patterns were observed between the RA of the OTUs and their reproducibility among technical replicates (data not shown).

Effects of variations in technical replicates on diversity estimations

To determine how such variations in technical replicates affect the estimations of microbial local diversity (α -diversity), the Shannon–Weaver index was calculated at levels of the technical replicates, biological replicates and treatments. Three-way ANOVA indicates that gene richness, functional gene diversity and evenness were all significantly different among technical replicates ($P < 0.05$) (Sup-

plementary Table S3), but most of them, if not all, were not significantly different at the biological replicate or treatment level. These results suggest that variations in technical replicates may not affect the estimation of α -diversity at biological replicate and/or treatment levels.

To understand whether variations in technical replicates affect the comparison of different microbial communities (that is, β -diversity), two popular similarity metrics, Sørensen's incidence-based and Bray-Curtis's abundance-based methods, are used. Their complements are used as metrics of β -diversity, which are widely used in many studies and range from 0 (when all OTUs/individuals are shared between two communities) to 1 (when no OTUs/individuals are shared). β -Diversity was calculated for each pair of microbial communities at the levels of technical replicates, biological replicates and treatments based on the combined OTU data from both forward and reverse primers (Table 1). The average β -diversities at the technical replicate level were 0.756 ± 0.042 based on the Sørensen index and 0.763 ± 0.049 based on the Bray-Curtis index without removing singletons (Table 1). These results indicated that the background noise in estimating β -diversity is quite high. However, in this case, one-way ANOVA revealed that the average β -diversity at both biological replicate and treatment levels based on both the Sørensen and the Bray-Curtis indices were all still significantly ($P < 0.05$) higher than those at the technical replicate level (Table 1). As expected, removing singletons significantly ($P < 0.05$) decreased β -diversity estimations at all three levels, but had more impact on the β -diversity estimation based on the Bray-Curtis index (Table 1).

Quantitation of the amplicon sequencing

To determine whether amplicon sequencing-based detection is quantitative, 10 pg gDNA from *S. oneidensis* MR-1 was spiked into PCR reactions

Table 2 Effects of data preprocessing on the significance (*P*-values) in resolving the differences of microbial communities between warming and unwarming^a

Primers	Not removing singletons			Removing singletons		
	Cut 0 ^b	Cut 1 ^c	Cut 2 ^d	Cut 0 ^b	Cut 1 ^c	Cut 2 ^d
Forward	0.55	0.05	0.03	0.44	0.07	0.02
Reverse	0.51	0.08	0.04	0.19	0.13	0.03
Forward and reverse	0.63	0.02	0.03	0.04	0.03	<0.01

Abbreviations: ANOSIM, analysis of similarity; MRPP, multiresponse permutation procedure.

^aThe values in the table are probability values (*P*), indicating significance with the Adonis method. Three statistical methods (MRPP, ANOSIM and Adonis) were used. However, Adonis is more sensitive; thus, only results using this method are presented.

^bNo OTUs were removed.

^cOTUs present only in 1 of the 14 tag sequence data sets of a treatment were removed.

^dOTUs present in 1 or 2 of the 14 tag sequence data sets of a treatment were removed.

Table 3 Effects of data preprocessing on the significance (*P*-values) in resolving the differences of microbial communities between clipping and unclipping^a

Primers	Not removing singletons			Removing singletons		
	Cut 0 ^b	Cut 1 ^c	Cut 2 ^d	Cut 0 ^b	Cut 1 ^c	Cut 2 ^d
Forward	0.6	0.1	0.03	0.59	0.07	0.04
Reverse	0.62	0.18	0.04	0.16	0.16	0.09
Forward and reverse	0.58	0.02	<0.01	0.06	0.03	<0.01

Abbreviations: ANOSIM, analysis of similarity; MRPP, multiresponse permutation procedure.

^aThe values in the table are probability values (*P*), indicating significance with the Adonis method. Three statistical methods (MRPP, ANOSIM and Adonis) were used. However, Adonis is more sensitive; thus, only results using this method are presented.

^bNo OTUs were removed.

^cOTUs present only in 1 of the 14 tag sequence data sets of a treatment were removed.

^dOTUs present only in 1 or 2 of the 14 tag sequence data sets of a treatment were removed.

containing 10 ng community DNA of each sample, and then PCR amplified and pyrosequenced. MR-1 has nine copies of the 16S rRNA gene, with 100% identity among them in the sequenced regions. Thus, only a single OTU from MR-1 would be obtained. Theoretically, 0.1% (wt/wt) sequences from the MR-1 genome would be detected in all of these samples. However, our experimental results indicated that the percentages of MR-1 OTUs detected varied substantially among different soil samples, ranging from 0.06% to 5.34% for the forward primer and 0.0% to 4.42% for the reverse primer. The difference between the maximum and minimum percentages of detected MR-1 sequences is ~66.7-fold for the forward primer. In contrast, no MR-1 sequence was detected in some tags for the reverse primer, whereas the maximum percentage of detected MR-1 sequences was 4.42% (Supplementary Table S4).

Improving data comparability

Various methods were used to improve data comparability, including increasing biological replicates, and removing singleton sequences and less-representative OTUs across biological replicates (see Supplementary text). Among three statistical methods (MRPP, ANOSIM and Adonis), which were used

to compare the differences of microbial community structure between warming and non-warming or clipping and non-clipping, Adonis seems to be the most sensitive (Tables 2 and 3, Supplementary Tables S5 and S6). Thus, only the results from Adonis are presented. The significance value (*P*) is presented in these tables and smaller *P*-values indicate better resolution. The following generalizations can be drawn. First, removing singleton sequences yielded better resolution (Tables 2 and 3). Removing less-representative OTUs is another option based on the results of biological replicates. Considerable differences in *P*-values were obtained by removing less-representative OTUs (one or two) across samples (Tables 2 and 3). In addition, generally only one-strand DNA sequences were obtained by amplicon sequencing, and hence, approximately half of the sequencing data are not used. In this study, both primers for PCR amplification were tagged, pyrosequenced and then assigned to OTUs. Thus, using sequencing reads from both forward and reverse primers leads to increased sequencing efforts. As expected, considerably better resolution was obtained using the combined sequencing data sets from both primers (Tables 2 and 3).

Despite several findings for improving data comparability, the real question is whether the effects of

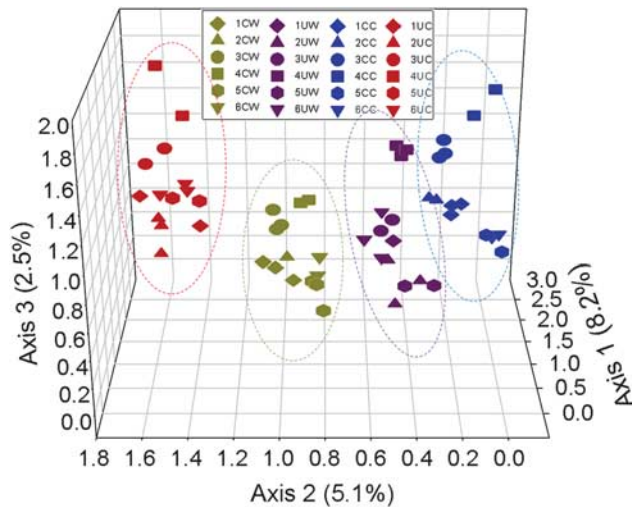


Figure 2 Detrended correspondence analysis (DCA) of 454 pyrosequencing data showing that warming and clipping significantly affected soil microbial community composition and structure. The relative abundance of all detected OTUs at 97% similarity cutoff in at least 4 of 14 tagged sequence data sets for each treatment were used for DCA analysis. Red dots are for unclipped, unwarmed control (UC), blue dots are for unwarmed, clipped (CC), dark purple dots are for unclipped, warmed (UW) and dark yellow dots are for clipped, warmed (CW). The effects of warming and clipping on soil microbial community composition and structure appeared to be well separated by the second axis.

warming and/or clipping on microbial communities can be detected based on such amplicon sequencing data. Detrended correspondence analysis showed that samples from warming and/or clipping plots clustered together and were well separated from control plots (Figure 2), suggesting that microbial community composition and structure were markedly different between warming and un-warming or between clipping and unclipping. To examine whether these observed differences are statistically significant, three complementary non-parametric multivariate statistical tests (ANOISM, Adonis and MRPP) were performed. The phylogenetic community structure based on the 16S rRNA gene was also significantly different between warming and un-warming, and between clipping and unclipping when more than five biological replicates were used, according to at least one of the three methods (Table 4). It is clear that increasing biological replicates provide better resolution in differentiating the treatment effects of warming or clipping. Taken together, these results indicated that the phylogenetic composition and structure of microbial communities under experimental warming or clipping were significantly different from those under control conditions.

Discussion

With the recent development and application of metagenomic technologies such as large-scale

Table 4 Significance tests of the effects of warming or clipping on the overall microbial community structure with three different statistical approaches

Data sets	Adonis ^a		ANOSIM ^b		MRPP ^c	
	F	P-value ^d	R	P-value ^d	δ	P-value ^d
<i>Warming vs un-warming</i>						
Six replicates	2.607	<0.01	0.519	<0.001	33.43	0.025
Five replicates	2.419	<0.01	0.5	0.009	33.12	0.052
Four replicates	1.960	0.04	0.427	0.059	32.02	0.107
Three replicates	1.797	0.06	0.482	0.179	30.82	0.220
<i>Clipping vs unclipping</i>						
Six replicates	2.243	<0.01	0.406	0.014	32.76	0.072
Five replicates	1.966	0.05	0.396	0.037	33.18	0.188
Four replicates	1.637	0.06	0.344	0.087	32	0.307
Three replicates	1.549	0.15	0.407	0.213	28.55	0.304

Abbreviations: ANOSIM, analysis of similarity; MRPP, multiresponse permutation procedure.

All three tests are non-parametric multivariate analyses based on dissimilarities among samples.

^aPermutational multivariate analysis of variance using distance matrices. Significance tests were performed by F-tests based on sequential sums of squares from permutations of the raw data.

^bANOSIM: Statistic *R* is based on the difference of mean ranks between groups and within groups. The significance of observed *R* is assessed by permuting the grouping vector to obtain the empirical distribution of *R* under the null model.

^cMRPP: Statistic δ is the overall weighted mean of within-group means of the pairwise dissimilarities among sampling units. The significance test is the fraction of permuted deltas that are less than the observed delta;

^dP-value of corresponding significance test.

The values in bold indicate the significant difference between the treatments (warming and clipping) and the control (at the levels of $P < 0.01$, or $P < 0.05$) by the tests.

high-throughput sequencing (Sogin *et al.*, 2006; Huber *et al.*, 2007; Hamady *et al.*, 2008), PhyloChip and GeoChip (He *et al.*, 2007, 2008, 2010a,b; Zhou *et al.*, 2008; Zhou, 2009; Vaishampayan *et al.*, 2010), spatial and temporal information on microbial community structure and functional activities can be rapidly obtained at a community-wide level. Metagenomic technologies allow scientists to address research questions, which could not be approached previously. High-throughput metagenomic technologies have greatly advanced microbiologists' capabilities for analyzing microbial communities in the environment and significantly shaped the field of microbial ecology. However, great caution is required when using high-throughput sequencing technologies for characterizing microbial community composition and structure as demonstrated in this study and in several others (Huse *et al.*, 2007; Gomez-Alvarez *et al.*, 2009; Engelbrektson *et al.*, 2010; Kunin *et al.*, 2010).

Despite several reports on the rigorous analyses of pyrosequencing error rates of 16S rRNA genes (Huse *et al.*, 2007; Kunin *et al.*, 2010), overestimation of rare biosphere communities (Huse *et al.*, 2010; Kunin *et al.*, 2010) and gene/taxon abundance (Gomez-Alvarez *et al.*, 2009), as well as PCR amplification conditions on estimating species

richness and evenness (Engelbrektson *et al.*, 2010), little is known about the technical reproducibility and quantitation of amplicon sequencing. Our results in this study demonstrated that the reproducibility of the detected OTUs among these technical replicates is quite low, with an average of $13.1\% \pm 1.5\%$ OTU overlap for the two tags and $5.9\% \pm 1.6\%$ OTU overlap for the three tags. Such low percentages of OTU overlap among technical replicates raise a concern of comparing the β -diversity of microbial communities across different samples for amplicon sequencing. Inherent high variations in technical replicates could be a significant problem for comparative studies (for example, β -diversity), but it could be less problematic for diversity discovery studies (for example, α -diversity). To make meaningful comparisons across different samples, variations among different samples must be larger than variations in technical replicates. Otherwise, it will be difficult to draw confident biological conclusions.

The high variations among technical replicates is most likely due to sampling artifacts associated with random sampling (Zhou *et al.*, 2008), as well as the performance of the technology *per se*. Many steps in the pyrotag-based sequencing analysis are associated with random sampling, including PCR amplification of target genes, ligation of amplified PCR products to sequencing adaptors for generating single-stranded DNA bound beads, emulsion and immobilization of beads, as well as bead deposition into wells on a PicoTiter Plate (454 Life Sciences). Given the vast complexity of a soil microbial community, the random sampling process could have more dramatic influence on estimating the difference of microbial communities across different samples (that is, β -diversity) (Zhou *et al.*, 2008). However, further theoretical analysis is required to support such speculation. In addition, based on rarefaction analysis, $<10\%$ of the microbial populations/genomes were sampled in each sampling event (that is, herein equivalent to sequencing a community with a tag). If microbial population abundance in the community is evenly distributed, theoretical values of 1% and 0.1% overlap would be expected for two and three sampling events, respectively. However, the actual percentages of OTU overlap were higher than the expected values (Figure 1), which could be explained by the facts that microbial populations could be not evenly distributed and dominant populations have a higher probability of being re-sampled. In addition, although this study addressed the random sampling problem of pyrosequencing, such problems should also exist in other similar sequencing technologies, such as Illumina and SOLiD sequencing platforms.

In general, only sequences from one strand are used for analysis with pyrotag sequencing. Therefore, about half of the available information is not used (Acosta-Martinez *et al.*, 2008; Hollister *et al.*, 2010). As demonstrated in this study, combining

sequences from both forward and reverse primers alleviates the problem of random sampling because it doubles the sampling effort. As demonstrated in this study, the statistical resolution is greatly improved when sequences from both ends are used for determining OTUs. To combine them together, an appropriate length of overlap of sequences from both ends is required. The 16S primers used in this study (F515, R907) seem to be an appropriate choice because they are highly conserved and their derived sequences have an appropriate length of overlap (123 bp) with the FLEX pyrosequencing technology. Various studies have demonstrated that the average length (240 bp) of DNA fragments generated by these primers should be sufficient to determine statistically significant differences among microbial genomes that reflect the most likely biology occurring in the environment (Edwards *et al.*, 2006; Sogin *et al.*, 2006).

Quantitative capability is a critical issue when sequencing-based technology is used for environmental and ecological studies. Using the control DNA spiked into PCR reactions, substantial variations in the control sequences were observed among the 56 tag sequencing, despite that various strategies were used to alleviate the amplification biases on quantitation, including combining several amplifications together and using less cycle numbers to avoid PCR product saturation. Therefore, amplicon sequencing is not quantitative. This observation is consistent with previous pyrotag sequencing studies (Engelbrektson *et al.*, 2010) and the general consensus that conventional PCR amplification is not quantitative (Suzuki and Giovannoni, 1996; Qiu *et al.*, 2001). Therefore, great caution is required when using the amplicon sequencing approach to make quantitative inferences of microbial community diversity in comparative studies.

One way to improve reproducibility and quantitation is to use biological replicates. As demonstrated in this study, increasing biological replicates greatly helps to differentiate the effects of warming or clipping on microbial communities and allows for a more rigorous statistical analysis, which is important for comparative studies. Notably, microarray-based technologies have significant advantages in minimizing/eliminating the sampling artifacts associated with random sampling processes (Zhou *et al.*, 2008); combining high-throughput sequencing with array-based technologies could provide robust measurements for comparative studies.

As demonstrated in this study, variations in technical replicates could lead to overestimating β -diversity of microbial communities. As such effects vary in experiments, depending on the complexity of microbial communities examined and sampling efforts, it is important to assess the effects of random sampling processes on estimating β -diversity for later appropriate data interpretation. Thus, we recommend that technical replicates (for example, more than three for each biological sample) should be considered in individual experi-

ments. These technical replicates will allow us to assess the background noise level in β -diversity estimation. With the most recent advanced sequencing technologies (such as Illumina HiSeq 2000), having several technical replicates for each biological sample should be not difficult to achieve. In addition, the three methods (MRPP, ANOSIM and Adonis) used in this study should be useful to assess the statistical significance of microbial community diversities under different conditions.

In conclusion, amplicon sequencing of the rRNA gene and other functional genes is rapidly becoming the method of choice for profiling microbial communities and has generated great insights into microbial community diversity. Although the amplicon sequencing-based detection approach is powerful, it suffers from several inherent drawbacks such as high sequencing errors and artifacts as demonstrated by several previous studies. This study revealed that the amplicon sequencing-based detection approach is not reproducible and quantitative with soil microbial communities, which was most likely caused by artifacts associated with random sampling processes. Such variations in technical replicates could have substantial effects on estimating β -diversity, but less on α -diversity. Therefore, great caution should be taken when amplicon sequencing-based detection is used for making quantitative inferences about the β -diversity of microbial communities. In addition, various strategies can be used to improve the comparability of amplicon sequencing data, including removing singleton sequences, adding sequences together from technical replicates, combining the sequences from both primers, removing less frequently encountered OTUs across biological replicates and increasing biological replicates. Consistent with many previous studies, application of the amplicon sequencing-based detection approach to analyze microbial communities from a long-term global change experiment revealed clear differences in the composition and structure of microbial communities between warming and non-warming or between clipping and non-clipping. These results suggest that if used appropriately, amplicon sequencing-based detection is useful for analyzing microbial community structure despite the fact that it has several inherent drawbacks, such as poor reproducibility and quantitation.

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